



The utility of near-infrared spectroscopy in the regression of low-frequency physiological noise from functional magnetic resonance imaging data

R.J. Cooper ^{a,*}, Louis Gagnon ^{a,b,c}, Daniel M. Goldenholz ^a, David A. Boas ^a, Douglas N. Greve ^a

^a Athinoula A. Martinos Center for Biomedical Imaging, Department of Radiology, Massachusetts General Hospital, Harvard Medical School, Charlestown, MA, USA

^b Harvard-MIT Division of Health Sciences and Technology, Cambridge, MA, USA

^c Department of Electrical Engineering and Computer Science, Massachusetts Institute of Technology, Cambridge, MA, USA

ARTICLE INFO

Article history:

Received 18 July 2011

Revised 6 October 2011

Accepted 9 November 2011

Available online 17 November 2011

Keywords:

Near-infrared spectroscopy

Functional magnetic resonance imaging

Low-frequency oscillations

Resting state

ABSTRACT

Near-infrared spectroscopy (NIRS) signals have been shown to correlate with resting-state BOLD-fMRI data across the whole brain volume, particularly at frequencies below 0.1 Hz. While the physiological origins of this correlation remain unclear, its existence may have a practical application in minimizing the background physiological noise present in BOLD-fMRI recordings. We performed simultaneous, resting-state fMRI and 28-channel NIRS in seven adult subjects in order to assess the utility of NIRS signals in the regression of physiological noise from fMRI data. We calculated the variance of the residual error in a general linear model of the baseline fMRI signal, and the reduction of this variance achieved by including NIRS signals in the model. In addition, we introduced a sequence of simulated hemodynamic response functions (HRFs) into the resting-state fMRI data of each subject in order to quantify the effectiveness of NIRS signals in optimizing the recovery of that HRF. For comparison, these calculations were also performed using a pulse and respiration RETROICOR model. Our results show that the use of 10 or more NIRS channels can reduce variance in the residual error by as much as 36% on average across the whole cortex. However the same number of low-pass filtered white noise regressors is shown to produce a reduction of 19%. The RETROICOR model obtained a variance reduction of 6.4%. Our HRF simulation showed that the mean-squared error (MSE) between the recovered and true HRFs is reduced by 21% on average when 10 NIRS channels are applied and by introducing an optimized time lag between the NIRS and fMRI time series, a single NIRS channel can provide an average MSE reduction of 14%. The RETROICOR model did not provide a significant change in MSE. By each of the metrics calculated, NIRS recording is shown to be of significant benefit to the regression of low-frequency physiological noise from fMRI data.

© 2011 Elsevier Inc. All rights reserved.

Introduction

Near-infrared spectroscopy (NIRS) uses variations in the absorption of near-infrared light to infer changes in the concentration of oxyhemoglobin (HbO) and deoxyhemoglobin (HbR) in the cerebral cortex (Franceschini and Boas, 2004; Gibson et al., 2005; Obrig and Villringer, 2003). The most common applications of NIRS employ a back-reflection geometry, such that both the source and detector of the near-infrared light are positioned on the same surface (Villringer et al., 1993). This geometry is usually necessary to ensure that an adequate amount of light can reach both the brain and then the detector, but it renders NIRS measurements highly sensitive to background hemodynamic oscillations in superficial tissues. These oscillations, due to heart rate, respiration and a variety of low-frequency effects, form the background physiological noise through which changes in cortical

HbO and HbR must be identified (Diamond et al., 2006; Lina et al., 2008; Prince et al., 2003).

The blood oxygen level dependent (BOLD) signal of functional magnetic resonance imaging (fMRI) occurs because of variations in the local concentration of the paramagnetic deoxyhemoglobin molecule (Huettel et al., 2004). As it is a hemodynamic measurement, BOLD-fMRI is also highly sensitive to background physiological oscillations and despite the autoregulatory actions of the brain, the effects of heart rate, respiration rate and low-frequency hemodynamic oscillations are often present in fMRI data (Hu et al., 1995).

A number of simultaneous NIRS and fMRI experiments have been performed in the study of functional activation (Huppert et al., 2006; Steinbrink et al., 2006; Strangman et al., 2002; Toronov et al., 2001). These studies have sought to investigate the relationship between the functional NIRS and BOLD signals in common regions of the brain, and have shown that there is a good correlation between the NIRS-recorded functional HbR response and the local BOLD signal. Recent studies have begun to investigate the relationship between NIRS and fMRI-BOLD during the resting state, i.e. when no stimulation paradigm is applied. Preliminary work by our group (Greve et al., 2009) and

* Corresponding author.

E-mail address: rcooper@nmr.mgh.harvard.edu (R.J. Cooper).

studies by Tong et al. (Tong and Frederick, 2010; Tong et al., 2011) have described a significant correlation between NIRS signals and BOLD-fMRI data across the whole brain volume during the resting state. A NIRS signal, obtained using sources and detectors placed on the scalp, appears to not only correlate with the BOLD signal present in the cortical voxels to which it is directly sensitive, but also to the BOLD signal of a large number of voxels spread throughout the brain. These resting-state correlations are particularly apparent at frequencies below 0.1 Hz (Greve et al., 2009; Tong and Frederick, 2010).

Tong and Frederick (2010) presented compelling evidence that the correlation which exists between resting-state NIRS and BOLD-fMRI is dominated by systemic oscillations in blood pressure which traverse the cerebral vasculature, rather than being suggestive of resting-state neural connectivity (Biswal et al., 1995). Systemic hemodynamic oscillations at 0.1 Hz and below are common to NIRS and BOLD-fMRI recordings and are thought to have two main sources; the variations in local vascular dilation known as vaso-motion (Gustafsson, 1993) and oscillations in arterial blood pressure known as Mayer waves (Julien, 2006). Although it was not explicitly proven, Tong and Frederick (2010) suggested that the component of the NIRS signal which correlates with BOLD-fMRI across the brain is primarily that derived from superficial tissues (i.e. the scalp) and not from the brain. While it is beyond doubt that NIRS signals contain a significant contribution from the cerebral cortex (and that this contribution will correlate with the local fMRI voxels), this contribution is invariably smaller than that of the superficial tissues (Boas et al., 2004; Obrig and Villringer, 2003). The dominance of superficial hemodynamic oscillations is partly what necessitates the detailed signal analysis and averaging common to functional NIRS experiments.

Irrelevant of its origin, the existence of a correlation between NIRS and fMRI-BOLD in the absence of stimulation has an obvious potential application: the minimization of physiological noise in fMRI experiments. The BOLD hemodynamic response function (HRF) typically provides a peak signal change of between 1 and 5% (Buxton et al., 1998; Friston et al., 2000), which can be significantly less than the level of physiological noise in the BOLD-fMRI signal. In order to maximize the efficiency with which the HRF can be recovered, fMRI experiments typically consist of carefully designed paradigms of stimulus repetition (Dale, 1999; Friston et al., 1999). The reliable identification of the physiological noise present in the BOLD signal allows for an increase in signal-to-noise ratio for a given experimental paradigm, or can allow the same signal-to-noise ratio to be achieved using fewer stimulations, minimizing the necessary duration of the fMRI scan. This goal is not insignificant, as fMRI experiments often require a subject to remain stationary in the scanner for periods of over an hour (Huettel et al., 2004). In addition to being expensive, this can cause serious discomfort and makes fMRI experiments less suitable for vulnerable subject groups.

A common approach to the minimization of fMRI physiological noise involves the distal measurement of hemodynamic oscillations and associated factors with a view to including these measurements in the linear model of the BOLD-fMRI signal. When applied in this way, such measurements are referred to as 'nuisance regressors'. Measurements of heart rate and heart rate variability (Bianciardi et al., 2009; Chang et al., 2009; Glover et al., 2000) respiration rate and respiration rate variability (Birn et al., 2006, 2008) and end-tidal carbon dioxide (Chang and Glover, 2009; Wise et al., 2004) have all been investigated as nuisance regressors. The most commonly applied nuisance regression technique is known as RETROspective Image CORrection (RETROICOR) (Glover et al., 2000). This approach uses distal measurements of pulse rate (via a pulse oximeter) and respiration (via a pneumatic chest belt) to enable an image-based correction of respiration and pulsatile oscillations in the BOLD signal.

For this study, we acquired simultaneous, resting-state fMRI and 28-channel NIRS in seven healthy adults in order to thoroughly assess the utility of NIRS signals as fMRI nuisance regressors. We calculate

the variance of the residual error in a general linear model of the baseline fMRI signal, and calculate the reduction in this variance that can be obtained by employing NIRS signals to regress out low-frequency physiological noise. To provide a more rigorous assessment, we simulate an event-related fMRI paradigm by introducing a series of synthetic HRFs into the resting-state fMRI data of our seven subjects in order to explicitly quantify the utility of NIRS in the recovery of the HRF using a finite impulse response (FIR) model (Burock and Dale, 2000). In both cases, we compare each NIRS model with a model containing both pulse and respiration RETROICOR regressors. We also assess the utility of a single NIRS channel in groups of voxels with different NIRS-fMRI correlation in order to quantify the success of a given NIRS regressor in terms of the baseline correlation between it and the BOLD-fMRI volume. Finally, we attempt to optimize the recovery of the HRF using a single NIRS channel by introducing a voxel-specific time lag between the NIRS and BOLD-fMRI time series.

Methods

Subjects and data acquisition

Seven healthy adult subjects were recruited to this study, which was approved by the Institutional Review Board of Massachusetts General Hospital. Each subject provided written, informed consent and underwent a single scanning session consisting of three resting-state runs lasting 300 s.

Near-infrared spectroscopy was performed using a bilaterally symmetric 28-channel probe and the TechEn CW6 continuous wave NIRS system operating at 690 and 830 nm, with a sample rate of 25 Hz (Franceschini et al., 2006). The NIRS probe covers a large proportion of the frontal and temporal lobes and is shown in position in Fig. 1.

All MRI data were acquired using a Siemens Tim Trio 3T system. Prior to functional imaging, an anatomical MRI was obtained using an MP-RAGE imaging sequence (TR = 2530 ms, TI = 1100 ms, TE = 3.44 ms). Functional MRI was performed using the Functional Biomedical Informatics Research Network protocol (fBIRN, www.birncommunity.org) (Greve et al., 2011) such that TR = 2 s, flip angle = 77°, TE = 30 ms, number of slices = 30, echo spacing = 500 ms, with posterior-anterior phase encoding and a bandwidth of 2290 Hz/pixel. The in-plane resolution was 3.43 mm with a slice thickness of 5 mm with a 1 mm skip and axial slices approximately aligned with the AC-PC line. Each functional run provided an fMRI time course of 150 data points. The scanner trigger pulse was acquired to allow the synchronization of the NIRS and fMRI recordings.

Cardiac and respiration waveforms were measured for all subjects using a pulse oximeter and either a pressure monitor or stress gauge attached to the chest with an elastic belt. Both acquisitions were sampled at 25 Hz.

NIRS pre-processing

The changes in optical density recorded at each of the 28 channels were converted to changes in concentration of HbO and HbR using the modified Beer-Lambert relationship, with a path length correction factor of 6 and a partial volume correction factor of 50 (Boas et al., 2004; Huppert et al., 2006). A NIRS channel was excluded from subsequent processing if the baseline detected intensity of either wavelength was below or above the acceptable range of the system (40 to 70 dB respectively) or if the channel was determined to be excessively noisy on visual inspection. Previous work has shown that NIRS-fMRI correlations are dominated by low-frequency components (Greve et al., 2009; Tong and Frederick, 2010). As a result our HbO and HbR waveforms were low-pass filtered at 0.1 Hz (Butterworth, 5th order) before being synchronized to the fMRI trigger pulse and down-sampled to the fMRI sample rate of 0.5 Hz. If no channels were

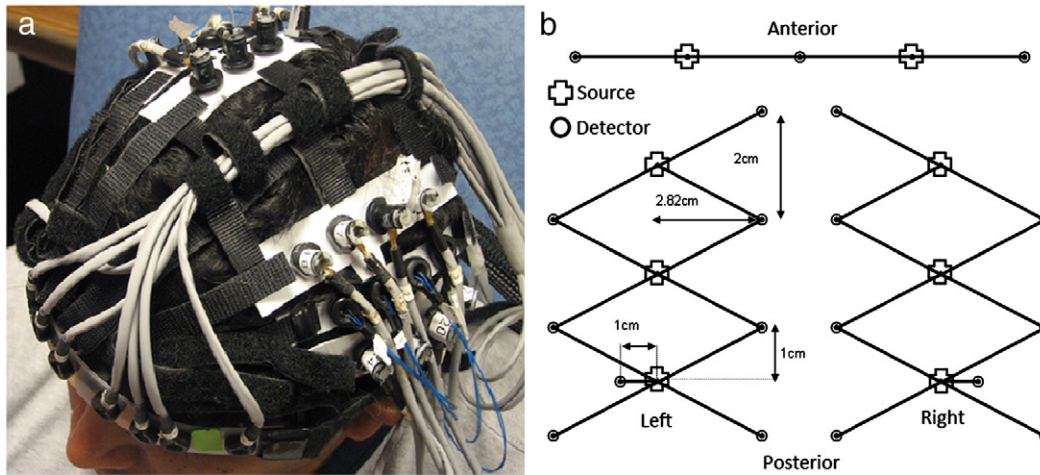


Fig. 1. The 28-channel NIRS array is shown in position on one subject in panel a. Panel b shows the schematic arrangement of the array.

rejected, this process would produce 56 time series (28 for HbO and 28 for HbR), each with 150 data points.

fMRI pre-processing

All fMRI scans were motion corrected to the middle time point using the AFNI 3dvolreg program (Cox and Jesmanowicz, 1999). Non-brain voxels were masked out using the FSL Brain Extraction Tool (Smith, 2002). Anatomical images were analyzed using FreeSurfer (surfer.nmr.mgh.harvard.edu) (Dale et al., 1999; Fischl et al., 1999) to construct models of the cortical surface of each individual. Each fMRI time series was smoothed using a 5 mm full-width at half-maximum Gaussian in the 3D volume. The middle time point of each run was registered to the subject's anatomical image using the FreeSurfer bregister program (Greve and Fischl, 2009) which allowed the subject-specific cortical mask to be mapped into the functional image space. All reported results are from cortical gray matter.

RETROICOR processing

Pulsatile and respiration regressors were created using the RETRO-spective Image CORrection (RETROICOR) approach (Glover et al., 2000). Pulse and respiration waveforms were modeled using a fundamental and first harmonic sine and cosine function to produce 4 regressors for pulse and 4 for respiration. These regressor waveforms were down-sampled to the fMRI acquisition rate on a slice-specific basis, so as to compensate for the finite delay between the acquisition of consecutive slices (Jones et al., 2008). The pulse oximetry data of one run in one subject was rejected due to a recording error. In this case only the 4 respiration RETROICOR regressors were used in further processing.

Linear models and error variance reduction

To assess the utility of NIRS regression of physiological noise, we constructed a number of linear models of the resting-state fMRI signal. All regression analyses were performed using FreeSurfer (surfer.nmr.mgh.harvard.edu) and bespoke Matlab (The Mathworks Inc.) software functions. The first linear model consisted of 3 regressors corresponding to the 0th, 1st and 2nd order temporal drift polynomials and a further 6 regressors constructed from motion correction parameters. This 'reference' model is used as a comparison throughout, and forms the basis of all other models.

A slice-specific RETROICOR model was computed by augmenting the reference model with all 4 pulse and all 4 respiration RETROICOR

waveforms for each slice as defined above. The regression was performed on a slice-specific basis and the results averaged across all cortical voxels.

The NIRS models were computed by adding one or more of the down-sampled NIRS signals to the reference model. The NIRS models are separated into 'HbO models' (those containing one or more HbO regressors), 'HbR models' (those containing one or more HbR regressors) and 'HbO & HbR models' (those containing both the HbO and HbR regressors from one or more NIRS channels). To provide a simple metric of the efficacy of NIRS modelling of resting-state fMRI time series, we computed the variance of the residual error of each model and compared it to that of the reference model. The larger the reduction in variance the better the model of the fMRI time series (a percentage variance reduction (PVR) of 100%, indicates that the model is able to remove all the variance in the fMRI time series unaccounted for by the reference model). The PVR achieved by the RETROICOR model was also calculated. A general linear model (GLM) of the fMRI time series (Y), with design matrix (X) has a residual error (ε) given by:

$$\varepsilon = Y - X\hat{\beta} \quad (1)$$

where $\hat{\beta}$ is the vector of parameter estimates calculated via:

$$\hat{\beta} = (X'X)^{-1}X'Y \quad (2)$$

The residual variance is given by:

$$\text{var}(\varepsilon) = \frac{\varepsilon^T \varepsilon}{d} \quad (3)$$

where d is an estimate of the model's degrees of freedom and is a function of the number of regressors included in the model. As the model is altered (for example by adding regressors to the design matrix, X), the residual variance ($\text{var}(\varepsilon)$) will change depending on how well the model accounts for data. Note that the variance does not necessarily decrease with the addition of regressors to the linear model.

HRF simulation

To further quantify the utility of NIRS regressors in the removal of physiological noise, and therefore the improved recovery of a functional response, we introduced a series of simulated HRFs into the resting-state fMRI time series of each data set. We produced a simple HRF using a single-gamma function (Fig. 7) with a peak latency of 6 s, down-sampled to match TR and scaled to provide a 2% peak increase in the

BOLD signal of each relevant voxel. A ‘stimulus paradigm’ of ten HRFs was introduced into each 300-second run in an event-related fashion. To increase the reliability of our results, 30 different stimulus paradigms were used in which each inter-HRF-interval was a randomly selected period between 2 and 24 s. For each stimulus paradigm, subject, run and linear model, the HRF was estimated using a finite impulse response approach. For comparison, this process was also performed for the RETROICOR model. The HRF recovered by each model was compared to the true HRF time series using two metrics: the mean-squared error (MSE) and Pearson’s correlation coefficient (R^2). Within the context of the GLM, the MSE is given by:

$$MSE = \frac{\text{var}(\varepsilon)}{\xi} \quad (4)$$

where the efficiency of the model (ξ) is:

$$\xi = \frac{1}{\text{trace}(C(X'X)^{-1}C')} \quad (5)$$

and C is the contrast matrix pertaining to the HRF (Dale, 1999). The value of the MSE achieved by a given model is thus dependent on both the residual variance and the efficiency of that model.

As the correlation between a given NIRS channel and the fMRI signal will vary significantly across the fMRI volume (Tong and Frederick, 2010), the selection of specific regions of interest in which to apply the simulated functional response so as to best test the use of NIRS signals as nuisance regressors is not trivial. To estimate the average effect of NIRS regression, the simulated HRFs were first added to (and recovered from) the BOLD time series of every cortical voxel, without any consideration for the correlation between the NIRS signals and the fMRI volume.

Selected voxels and optimized time lag

This simulation was repeated for models containing only a single NIRS regressor in groups of 100 voxels selected on the basis of the value of the correlation between the fMRI time series and each NIRS regressor. This process was performed in order to quantify the success of a given NIRS regressor with respect to the value of the baseline correlation between it and the BOLD-fMRI volume. The baseline correlation is denoted R_{base}^2 to distinguish it from the correlation between synthetic and recovered HRFs (R^2). We calculated R_{base}^2 using the fMRI time series prior to the introduction of the simulated HRF in order to minimize computational expense. Three groups of 100 voxels, with R_{base}^2 in the range 0–0.1, 0.1–0.2 and 0.2–0.3, were selected for each subject, run and HbO or HbR signal.

The results of Tong and Frederick (2010) suggest that the NIRS–fMRI correlation traverses the cerebral vasculature in a wave-like pattern. In an attempt to take advantage of this phenomenon, the HRF simulation was also performed across the whole cortex using models containing a single NIRS regressor, but with a voxel-specific time lag introduced between that NIRS regressor and the fMRI time series. Because the NIRS recording time for each run exceeded that of the fMRI (NIRS recording began ~30 s prior to the fMRI sequence and continued for ~30 s after) we were able to shift the NIRS signals in time relative to the fMRI recording. We produced 20 different time lags, from –10 to +10 s in 1-second intervals, relative to the fMRI time series for every HbO and HbR signal. For every cortical voxel, HbO signal, HbR signal and for every run, we were able to compute the time lag which optimized the baseline correlation (R_{base}^2). A voxel and NIRS signal specific linear model was then used to recover the simulated HRF at each voxel using this optimum time lag, and the results were averaged across the cortex.

Synthetic NIRS signals

Near-infrared spectroscopy signals will always contain some noise contribution from non-physiological sources. When the NIRS signals are low-pass filtered, this noise is also low-pass filtered. When such waveforms are used in the general linear model, it is possible that they could provide a good model of resting-state fMRI oscillations simply because the constituent noise spans the frequency space below 0.1 Hz. The use of NIRS regressors could therefore produce a significant reduction in fMRI noise that is not related to the physiological measurements of HbO and HbR. To account for this possibility, our calculations of variance reduction and our HRF simulations were all repeated using synthetic NIRS data. These waveforms were obtained by producing two random vectors and passing them through the entire processing stream, including the Beer–Lambert transformation and low-pass filtering. This is effectively equivalent to repeating all our processing under the null hypothesis that there is no relationship between HbO, HbR and the BOLD signal.

Results

The average proportion of the cortex found to be significantly correlated with a given NIRS regressor (using a two-tailed, voxel-wise t -test, $p < 0.01$) was 17.8% for HbR signals and 16.8% for HbO signals, with subject means ranging from 5 to 22%. Subject 6 showed very little correlation despite there being no obvious issues with either the NIRS or fMRI recordings. Fig. 2 shows an example of a significance map for a single, well-correlated HbO regressor. The average proportion of the cortex significantly correlated with the synthetic NIRS signals was 3.7%.

Whole-cortex variance reduction

The percentage variance reduction (PVR) was computed for the HbO, HbR, HbO & HbR, RETROICOR and synthetic NIRS models. The results of this process are shown in Fig. 3a as a function of the number of NIRS channels (n) in each linear model.

Each data point in Fig. 3a corresponds to the mean PVR achieved over the whole cortex and averaged over all runs and either every possible combination of n channels or 28 random combinations of n channels, whichever is smaller. For example, the HbO data point corresponding to 1 NIRS channel is the average PVR obtained across every possible model containing a single HbO regressor (if no channels were excluded, there would be 28 possible models), for each subject and run. For the HbO data point corresponding to 2 NIRS channels, 28 pairs of non-excluded HbO regressors were randomly selected, and the average PVR achieved by those pairs of regressors is presented. The upper limit on the number of random combinations was set at 28, as it was computationally prohibitive to use every possible model in most cases (for example, 28 choose 2 = 378 possible models). This process was performed in order to minimize the sensitivity of the results to channel selection. This same process was applied to obtain the HbR model PVR results in Fig. 3. For the HbO & HbR models, 1 NIRS channel corresponds to 2 regressors (the HbO and HbR regressors corresponding to each single NIRS channel), 2 NIRS channels corresponds to 4 regressors and so on. This randomized selection approach was also applied for the whole-cortex HRF simulation results below.

As shown in Fig. 3a, the PVR increases with the addition of NIRS regressors for all models until 15 NIRS channels are applied, at which point the PVR is equal to 36% for the HbO & HbR model. The PVR is significantly larger than zero for all models (one-tail t -test, $p < 0.01$). The addition of the slice-specific respiration and pulse RETROICOR regressors to the reference model also produces a significant reduction in variance, averaging 6.4%. The PVR achieved by the HbO & HbR model becomes significantly larger than that of the RETROICOR model when 2 NIRS channels are applied (one-tail, paired t -test, $p < 0.01$). It is worth noting that the use of 8 synthetic NIRS regressors

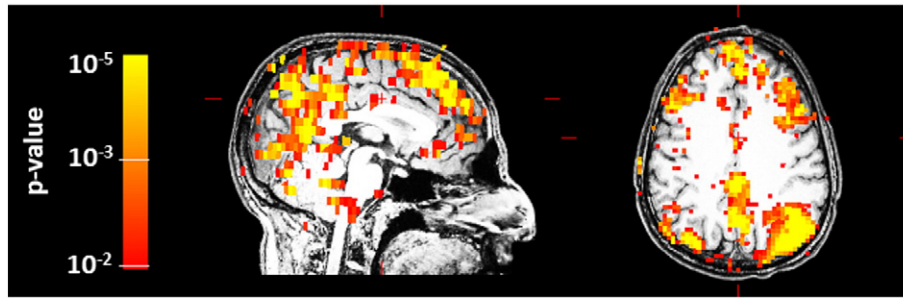


Fig. 2. An example of the significance map for a single subject, run and NIRS-HbO regressor. Note that significantly correlated voxels are spread throughout the cortex as well as in deeper regions of the brain.

(the same number of regressors used in the RETROICOR model) produces an average PVR of 4.7%.

Fig. 3b shows the corrected PVR, achieved by subtraction of the variance reduction calculated using the synthetic NIRS waveforms. This provides a lower-bound estimate of the average PVR attributable to each NIRS model. A peak value of 17.0% is achieved for the HbO & HbR model incorporating 9 NIRS channels. The PVR remains significantly larger than zero for all models after subtraction of the synthetic NIRS results (one-tail t -test, $p < 0.01$).

Whole-cortex HRF simulation

Fig. 4a shows the average mean-squared error (MSE) between the synthetic and recovered HRFs for HbO, HbR, HbO & HbR, RETROICOR and the synthetic HbO & HbR models (synthetic HbO and synthetic HbR models were also calculated but for simplicity are not displayed).

These data are normalized to the value obtained using the reference model (i.e. that without NIRS regressors) and are shown as a function of the number of NIRS channels applied (n). For each NIRS model, either all possible combinations, or 28 combinations of n NIRS channels were randomly selected and the results provided are an average of all combinations, as described above for Fig. 3. The MSE decreases with the addition of NIRS regressors for HbO, HbR and HbO & HbR models. This decrease reaches significance (one-tail paired t -test, $p < 0.01$) with the use of a single NIRS channel for the HbO & HbR model and with the use of 2 NIRS channels for both the HbO and HbR models. The minimum, equal to reduction of 21%, is found for the HbO & HbR model when 10 NIRS channels are applied.

The slice-specific RETROICOR results are highly variable but on average the RETROICOR model fails to reduce the MSE between the synthetic and recovered HRFs, and results in an average increase in MSE of 0.1% (standard deviation of 14%) compared to the reference model.

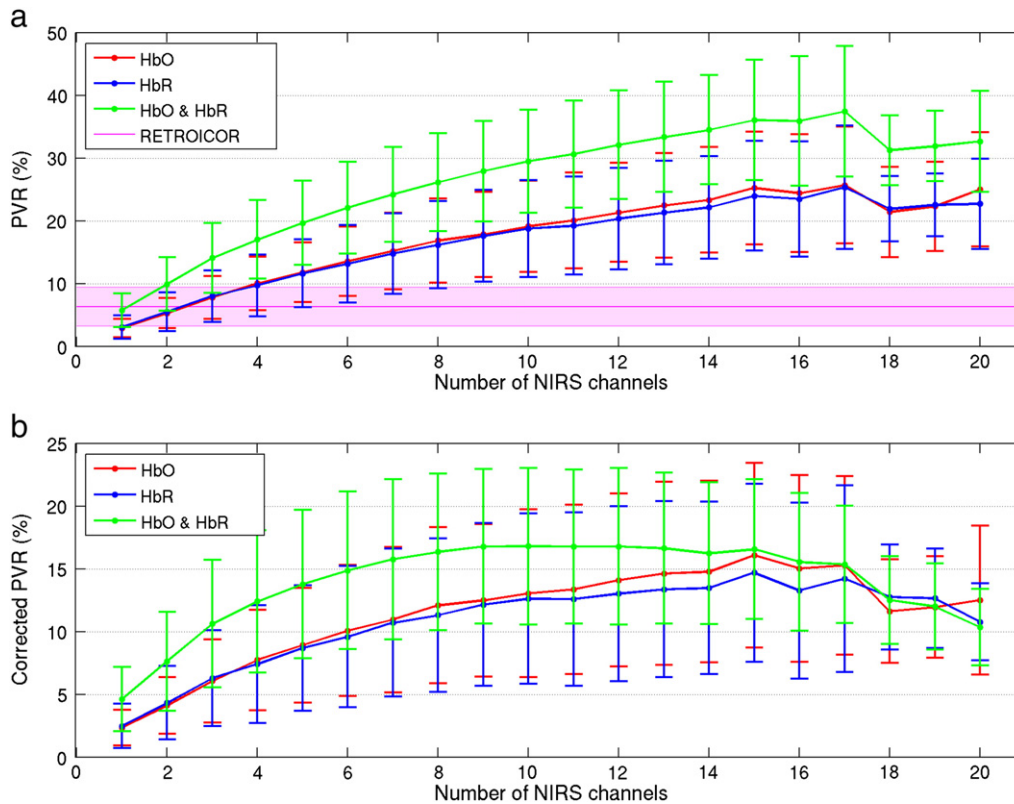


Fig. 3. The percentage variance reduction obtained by the HbO, HbR and HbO & HbR models as a function of the number of NIRS channels incorporated into each model. Values of PVR are obtained in comparison to the reference model. The corresponding results for the pulse and respiration RETROICOR model are also included. Panel b shows the corrected PVR values, which were obtained by subtracting the PVR achieved using synthetic NIRS regressors from that of the real NIRS regressors. These corrected results represent the extent to which the variance of the BOLD-fMRI signal can be modeled by the physiological content of the NIRS data. In each figure the error bars (and shaded region for the RETROICOR model) represent the standard deviation across all runs.

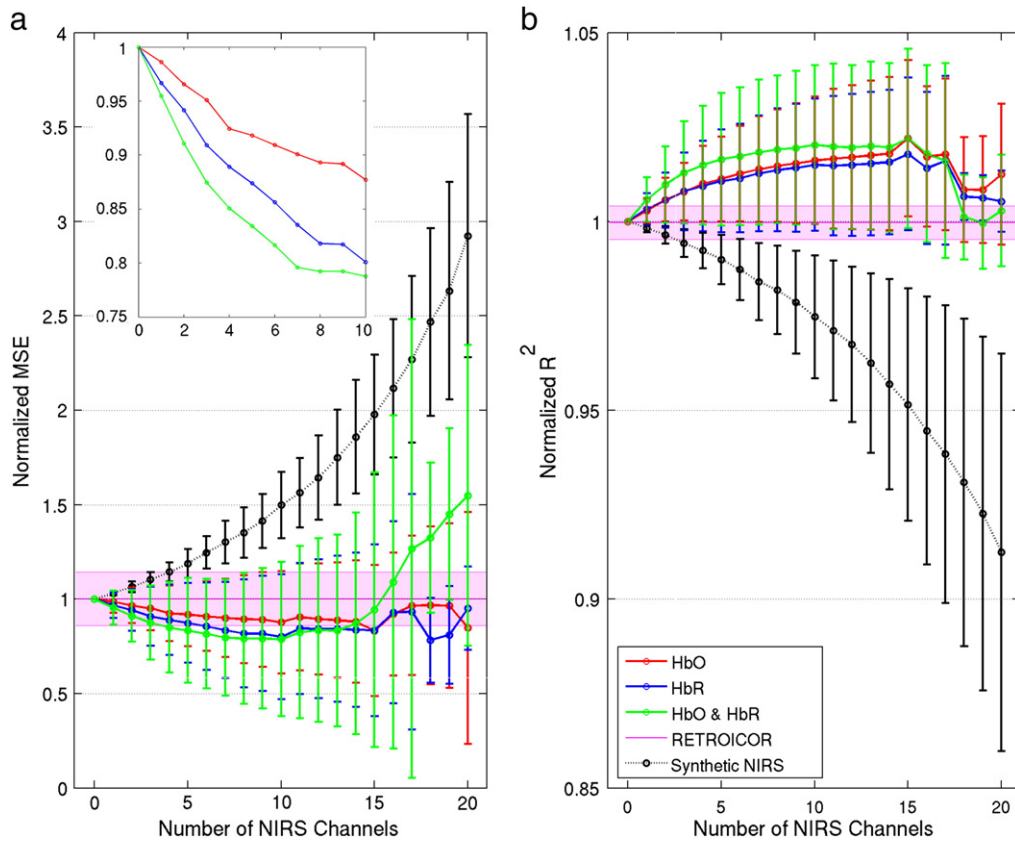


Fig. 4. The result of the HRF simulation applied across all cortical voxels for HbO, HbR, HbO & HbR and synthetic NIRS models as a function of the number of NIRS channels applied. The corresponding results for the pulse and respiration RETROICOR model are also included. Fig. 4a shows the mean-squared error (MSE) between the synthetic and recovered HRFs, normalized to the value obtained by the reference model. For clarity, the normalized MSE for up to ten NIRS channels are shown on a larger scale on the inserted axes. Panel b shows Pearson's correlation coefficient (R^2) between the synthetic and recovered HRFs. The mean changes in MSE and R^2 for the RETROICOR model compared to the reference model are negligible. In each figure the error bars (and shaded region for the RETROICOR model) represent the standard deviation across all runs.

Fig. 4b shows the corresponding results for the correlation coefficient (R^2) between the synthetic and recovered HRFs. Again, the data are normalized to the reference model. The value of R^2 between the synthetic and recovered HRFs follows a similar pattern to that observed in the MSE results; an improvement is observed with the number of NIRS channels applied for all models up to 10 NIRS channels, at which point the HbO & HbR model produced an increase in R^2 of 2.0%. This increase in R^2 above the reference model achieves significance for all NIRS models when a single NIRS channel is applied (one-tail paired t -test, $p < 0.01$). The RETROICOR model also fails, on average, to produce an improvement in R^2 , producing a mean decrease of 0.04% (standard deviation 0.4%).

The equivalent results calculated using the synthetic NIRS data show an approximately exponential increase in MSE and decrease in R^2 as synthetic regressors are added. This detrimental effect reaches significance for both MSE and R^2 metrics for all synthetic NIRS models using a single NIRS channel.

Selected voxels and optimized time lag

Figs. 5a–c show scatter plots of the MSE for single-regressor HbO and HbR models plotted against the corresponding MSE of the reference model. Data points on the 45° line correspond to instances where the NIRS regressor model has had no effect on the MSE. These simulations were performed in 100 voxels specific to each subject, run and HbO or HbR regressor such that R_{base}^2 lies between 0–0.1, 0.1–0.2 and 0.2–0.3 for Figs. 5a–c respectively. This process was not repeated for the synthetic NIRS signals because their correlation with the fMRI data was insufficient.

For voxels in the 0–0.1 baseline correlation band (Fig. 5a), NIRS regression provides no significant change in MSE for either the HbO or HbR models when compared to the baseline model (two-tailed t -test, $p = 0.53$). Both the 0.1–0.2 and 0.2–0.3 baseline correlation bands (Figs. 5b and c) show a significant decrease in MSE for both HbO and HbR models ($p < 0.01$). Fig. 5d shows the mean percentage change in MSE for each of these three bands. Figs. 6a–d are equivalent to Figs. 5a–d but present the value of the correlation coefficient (R^2) between the synthetic and recovered HRF instead of the MSE. Fig. 6d shows that the mean value of R^2 increases as a function of the baseline correlation R_{base}^2 . Fig. 7 provides one example of the effect of HbO (Fig. 7a) and HbR (Fig. 7b) regressors in well-correlated voxels ($0.2 < R_{\text{base}}^2 < 0.3$), for a single run and subject.

Fig. 8 shows the results of the HRF simulation where the optimum time lag has been introduced between each NIRS-HbO or HbR signal and the fMRI time series of each voxel. Figs. 8a and b respectively show the MSE and R^2 between the synthetic and recovered HRFs, normalized to the values obtained by the reference model. In each case the result of the comparable zero-time lag models (equivalent to the first data points of Figs. 4a and b) is provided for comparison. The use of the optimum time lag causes a significant reduction in MSE (one-tailed paired t -test, $p < 0.01$) for all models compared to the zero-time lag equivalent, including the synthetic NIRS model. The reduction in MSE obtained for the optimum time-lag HbO & HbR model is 13.6% compared to 6.3% for zero time lag. Correspondingly, there is an increase in R^2 when the optimum time lag regressors are applied, but this does not quite reach significance for the HbO ($p = 0.011$), HbR ($p = 0.014$) or HbO & HbR ($p = 0.10$) models. The improvement is very significant for the synthetic NIRS data ($p < 10^{-6}$). For both metrics, the use of

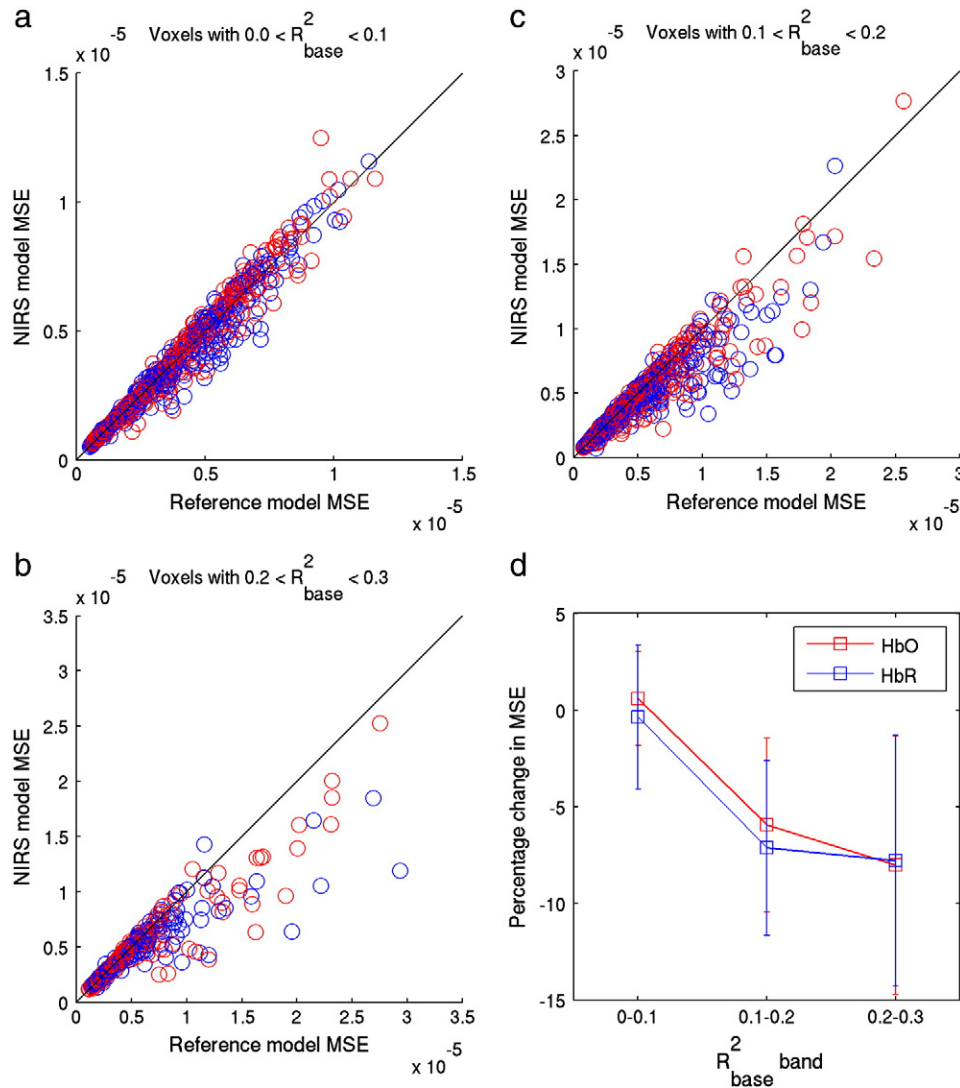


Fig. 5. The single-regressor NIRS model MSE plotted against the reference model MSE for 100 voxels selected using the value of the baseline NIRS-fMRI correlation for each voxel. The data is separated into three bands on the basis of R^2_{base} . The average reduction in MSE for all data points is shown in panel d. The MSE between the synthetic and recovered HRFs decreases as a function of R^2_{base} . The error bars in panel d show the standard deviation across all data points of panels a–c.

optimum time lag produces a greater improvement for HbO regressors than for HbR regressors.

Discussion

Our results confirm that low-frequency oscillations present in near-infrared spectroscopy data correlate significantly with a large proportion of BOLD-fMRI voxels, spread throughout the cortex. At frequencies below 0.1 Hz, NIRS signals are subject to a range of physiological oscillations. These include the spontaneous changes in vascular dilation referred to as vaso-motion (Gustafsson, 1993) and the global oscillations in arterial blood pressure known as Mayer waves (Julien, 2006). As the effects of vaso-motion tend to be location-specific, the systemic oscillations caused by Mayer waves affecting both the scalp-dominated resting-state NIRS signal and BOLD-fMRI voxels across the brain seem the most likely source of this correlation.

Whole-cortex PVR and HRF simulation

The utility of NIRS signals in the regression of physiological noise can be established in a variety of ways. A simple assessment of the efficacy of a given linear model is the variance of that model's residual error. Our

results indicate that the inclusion of NIRS signals in the model can reduce this variance by as much as 36%. To achieve this, a large number of NIRS channels (> 15) are required. NIRS regression produces an average PVR of 6.0% for a single NIRS channel, comparable to that achieved by the slice-specific RETROICOR approach (6.4%), (Fig. 3a).

As is clear from Fig. 3, a significant proportion of the variance reduction achieved by NIRS regression is not due to the physiological information present in the NIRS data, but can equally be achieved by the regression of random vectors occupying the same frequency space. However, even when this effect is taken into account, an average PVR of 17% can still be achieved by the HbO & HbR model (Fig. 3b). These corrected PVR results suggest that applying 10 NIRS channels provides the best model of physiological noise in the BOLD signal. The value of 17% represents a lower-bound estimate on the average PVR which can be obtained by a model containing 10 or more NIRS channels.

The results of our explicit simulation of an event-related BOLD-fMRI experiment confirm that NIRS can be used to improve the recovery of the HRF time course. There is a significant improvement in both the mean-squared error (MSE) and the correlation coefficient (R^2) between the synthetic and recovered HRFs when NIRS regressors are applied. Note that both the HbO and HbR models provide an improvement in

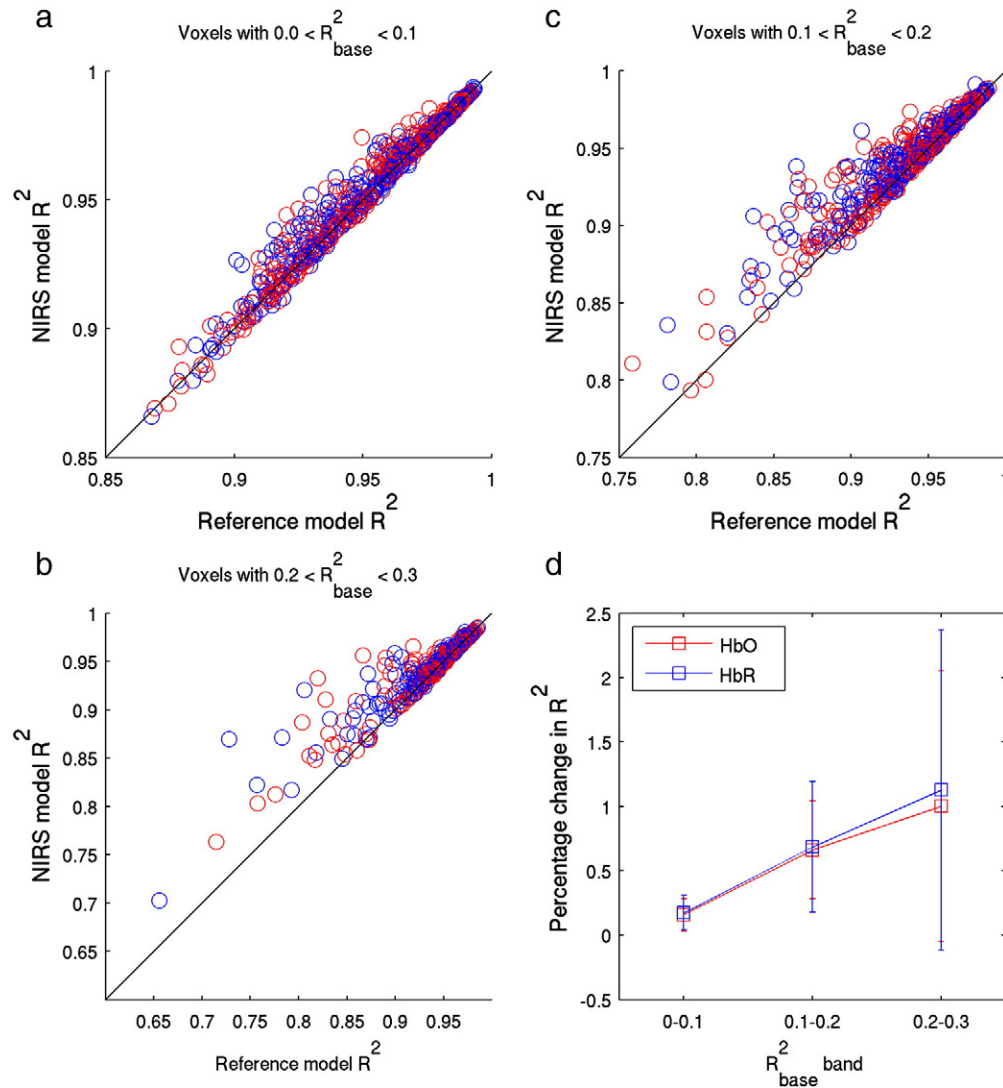


Fig. 6. The single-regressor NIRS model R^2 plotted against the reference model R^2 for 100 voxels selected using the value of the baseline NIRS–fMRI correlation for each voxel. The data is separated into three bands on the basis of R_{base}^2 . The average increase in R^2 for all data points is shown in panel d. The value of R^2 between the synthetic and recovered HRFs increases as a function of R_{base}^2 . The error bars in panel d show the standard deviation across all data points of panels a–c.

MSE and R^2 and that the HbO & HbR model is consistently better than either single-chromophore model. This may suggest that the HbO and HbR signals provide independent contributions to the model of resting-state fMRI fluctuations, but could also be the result of spatial averaging if the HbO and HbR signals are correlated with different regions of the cortex.

Fig. 4 shows that recovery of the HRF improves even if regressors are applied in a ‘blind’ fashion; across the whole cortex and without consideration for the baseline NIRS–fMRI correlation. The minimum MSE and maximum R^2 are achieved across the whole cortex when approximately 10 NIRS channels are included in the linear model. The MSE does not monotonically decrease (as the PVR increases in Fig. 3a) because the MSE is dependent on both variance and efficiency. Although adding NIRS regressors decreases the residual variance, it can also decrease the model’s efficiency. As the number of NIRS channels increases beyond 10, the decrease in efficiency overcomes the decrease in residual variance, and the MSE begins to increase.

As stated previously, we believe that the majority of the low-frequency contribution to the resting-state NIRS signal (and therefore to the NIRS–fMRI correlation) originates in the superficial layers of the head. As a result, the acquisition of NIRS signals suitable for regression could be performed using channels which only sample superficial tissues. Because of the improved signal-to-noise ratio associated

with short separation channels, superficial NIRS acquisition is often significantly easier than standard NIRS. Obtaining multi-channel, superficial NIRS during fMRI experiments is therefore entirely feasible, and may be particularly useful in experiments which require precise recovery of the HRF time course and where the number of stimuli or the scanning duration are limited. Examples of such cases include the study of the hemodynamic response to inter-ictal epileptic activity with fMRI (Bénar et al., 2002) or the computation of the cerebral metabolic rate of oxygen from combined NIRS–fMRI (Huppert et al., 2009). It is important to note that the HRF simulation data presented here are the average results obtained from 30 pseudo-random event-related paradigm designs. It was necessary to use a large number of different designs in order to produce a robust result, which suggests that the efficacy of NIRS regression may be sensitive to experimental design. An investigation of the utility of NIRS in the regression of fMRI noise for different experimental paradigms (including block designs) will therefore be necessary to establish which fMRI experiments will benefit from simultaneous NIRS recording.

Selected voxels and optimized time lag

The results summarized in Figs. 5 and 6 provide a measure of the importance of the baseline correlation (R_{base}^2) to the success of each

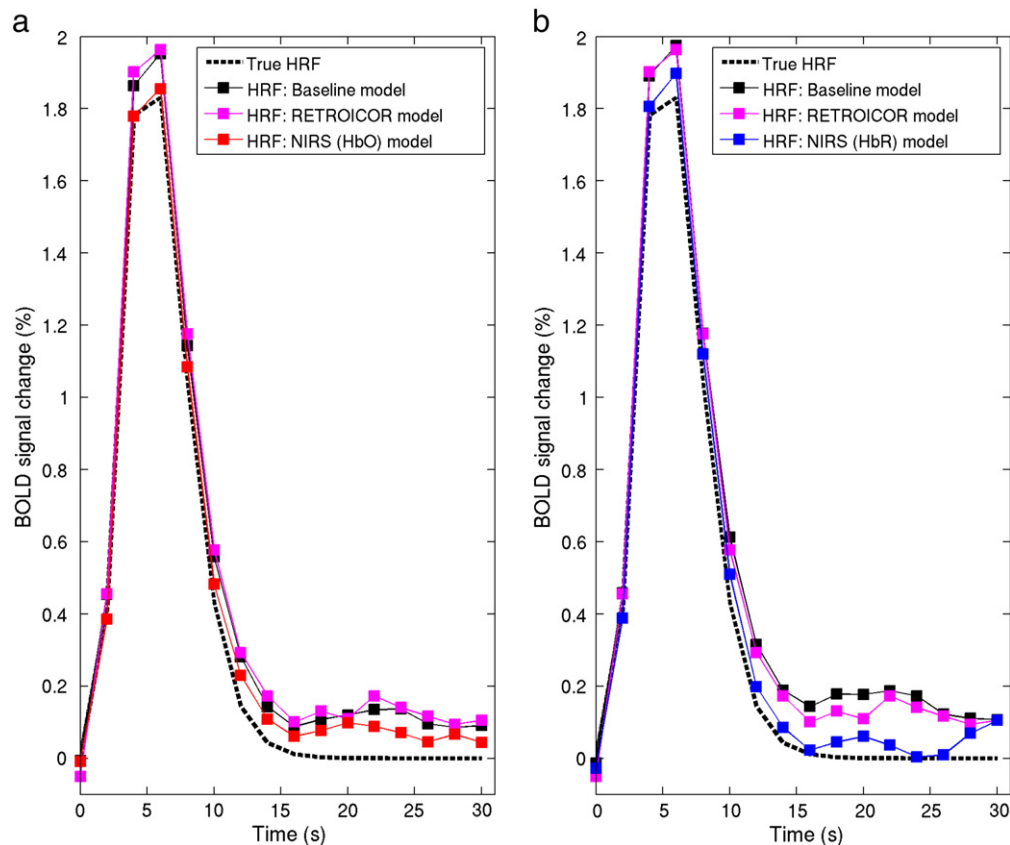


Fig. 7. An example of the HRFs recovered using the reference model, pulse and respiration RETROICOR model and by single-regressor HbO (panel a) and HbR (panel b) models for a single subject, run and NIRS channel for voxels well correlated each NIRS regressor ($0.2 < R_{\text{base}}^2 < 0.3$). The true, single-gamma HRF is also shown in both panels.

NIRS model. Although it is difficult to obtain the statistical power required to provide exact quantification of this relationship (due to variation in BOLD noise in different groups of voxels for example), the use of a single NIRS regressor in voxels where $R_{\text{base}}^2 > 0.1$ will, on average, produce a significant improvement in HRF recovery.

It is also important to note that even in cases of low baseline correlation (Figs. 5a and 6a), the use of a single NIRS regressor does not, on average, worsen the recovery of the HRF. This is consistent with Fig. 4, which shows that a significant improvement is observed even when NIRS regression is applied across the whole cortex, which must include regions and channels which exhibit a poor NIRS–fMRI correlation. This does not appear to be the case for the synthetic NIRS signals, as the whole-cortex simulation shows that HRF recovery worsens significantly with the use of a single synthetic regressor.

The results of employing the optimum time lag between each NIRS and fMRI time series show a significant decrease in MSE and a modest increase in R^2 for HbO, HbR and HbO & HbR models. A comparable improvement in HRF recovery is seen for the synthetic NIRS regressors (Fig. 8). An improvement in HRF recovery is clearly expected because by selecting the optimum time lag we are maximizing the value of the baseline correlation between each NIRS signal and the fMRI data. The fact that the synthetic NIRS regressors (even when optimized for time lag) do not provide the reduction in MSE achieved by the real NIRS signals is further evidence that low-frequency physiological oscillations are common to both fMRI and NIRS recordings. These results are also consistent with the hypothesis that the NIRS–fMRI correlation follows a low-frequency wave pattern throughout the cerebral vasculature, but the improvement this affords to HRF recovery is relatively small; comparable to the addition of a second NIRS channel.

The results of our simulations in pre-selected voxels, and the results of our time-lag optimization are both reliant on a calculation of the baseline correlation (R_{base}^2) between each NIRS signal and the fMRI data *prior* to the introduction of the simulated HRF series. The improvement in the recovery of the HRF associated with a higher baseline correlation is therefore to be expected. However, the introduction of the HRF series into the fMRI data produced a negligible change in R_{base}^2 ; 0.0058 for HbO and 0.0059 for HbR signals. It is therefore reasonable to suggest that NIRS regression could be optimized in a real fMRI study by maximizing R_{base}^2 , as calculated using the fMRI data *during* the stimulation period, with little risk of removing the HRF itself. This would obviously require that the NIRS channels are arranged so as not to sample the activated regions of the cortex. Alternatively, R_{base}^2 could be calculated for a period of NIRS–fMRI data where no stimulation occurs (i.e. during a rest period), but this assumes that the scale and spatial distribution of the NIRS–fMRI correlation will remain constant in time, the veracity of which has yet to be determined.

Conclusions

We have demonstrated that NIRS recordings can be employed to reduce the variance of the residual error in a general linear model of the resting-state BOLD–fMRI. Our results show that the use of a single NIRS channel will reduce variance in the residual error by an average of 6.0%, comparable to that achieved by RETROICOR methods. The use of 10 NIRS channels can reduce variance by as much as 36% on average across the whole cortex, though the same number of low-pass filtered white noise regressors can achieve a PVR of 19%. We have also explicitly shown that the use of even a single NIRS channel can significantly improve the recovery of a simulated hemodynamic response function

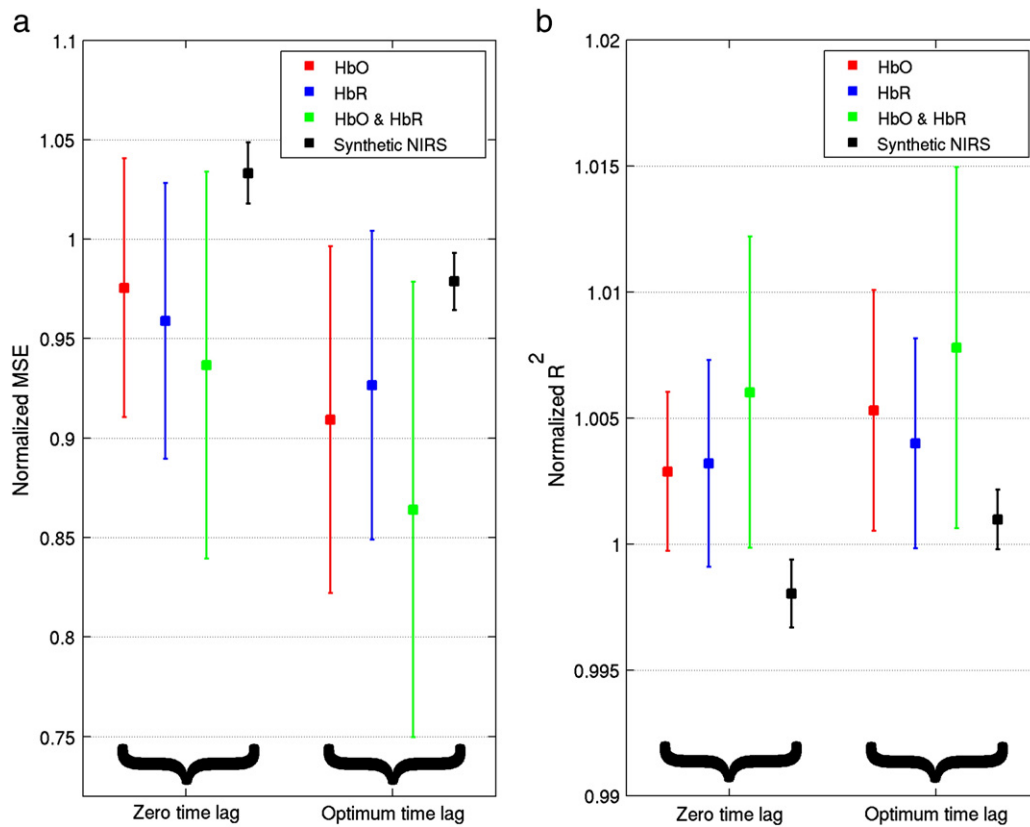


Fig. 8. The values of the MSE and R^2 between the synthetic and recovered HRFs for the zero time lag and optimum time lag, single-channel HbO, HbR, HbO & HbR and synthetic NIRS models, are normalized to the equivalent values of the reference model. A significant (two-tailed paired t -test, $p < 0.01$) improvement is apparent for both metrics and all models, though the improvement is more distinct for HbO models than for HbR models. In each case the error bars represent the standard deviation across all runs.

from BOLD-fMRI data, compared to standard fMRI and RETROICOR approaches. The MSE between the recovered and synthetic HRFs can be reduced by as much as 21% when 10 NIRS channels are applied. While the exact physiological origins and the inter-subject (and intra-subject) variability of NIRS-fMRI correlations must be examined further, it is clear that they have an important application in the enhancement of fMRI experiments.

Acknowledgments

This work was supported by NIH grants P41-RR14075 and R01-EB006385. L Gagnon is supported by the Fond Quebecois de Recherche sur la Nature et les Technologies.

References

Bénar, C.-G., Gross, D.W., Wang, Y., Petre, V., Pike, B., Dubeau, F., Gotman, J., 2002. The BOLD response to interictal epileptiform discharges. *Neuroimage* 17, 1182–1192.

Bianciardi, M., Fukunaga, M., van Gelderen, P., Horowitz, S.G., de Zwart, J.A., Duyn, J.H., 2009. Modulation of spontaneous fMRI activity in human visual cortex by behavioral state. *Neuroimage* 45, 160–168.

Birn, R.M., Diamond, J.B., Smith, M.A., Bandettini, P.A., 2006. Separating respiratory-variation-related fluctuations from neuronal-activity-related fluctuations in fMRI. *Neuroimage* 31, 1536–1548.

Birn, R.M., Smith, M.A., Jones, T.B., Bandettini, P.A., 2008. The respiration response function: the temporal dynamics of fMRI signal fluctuations related to changes in respiration. *Neuroimage* 40, 644–654.

Biswal, B., Yetkin, F.Z., Haughton, V.M., Hyde, J.S., 1995. Functional connectivity in the motor cortex of resting human brain using echo-planar MRI. *Magn. Reson. Med.* 34, 537–541.

Boas, D.A., Dale, A.M., Franceschini, M.A., 2004. Diffuse optical imaging of brain activation: approaches to optimizing image sensitivity, resolution, and accuracy. *Neuroimage* 23, S275–S288.

Burock, M.A., Dale, A.M., 2000. Estimation and detection of event-related fMRI signals with temporally correlated noise: a statistically efficient and unbiased approach. *Hum. Brain Mapp.* 11, 249–260.

Buxton, R.B., Wong, E.C., Frank, L.R., 1998. Dynamics of blood flow and oxygenation changes during brain activation: the balloon model. *Magn. Reson. Med.* 39, 855–864.

Chang, C., Glover, G.H., 2009. Relationship between respiration, end-tidal CO₂, and BOLD signals in resting-state fMRI. *Neuroimage* 47, 1381–1393.

Chang, C., Cunningham, J.P., Glover, G.H., 2009. Influence of heart rate on the BOLD signal: the cardiac response function. *Neuroimage* 44, 857–869.

Cox, R.W., Jesmanowicz, A., 1999. Real-time 3D image registration for functional MRI. *Magn. Reson. Med.* 42, 1014–1018.

Dale, A.M., 1999. Optimal experimental design for event-related fMRI. *Hum. Brain Mapp.* 8, 109–114.

Dale, A.M., Fischl, B., Sereno, M.I., 1999. Cortical surface-based analysis: I. Segmentation and surface reconstruction. *Neuroimage* 9, 179–194.

Diamond, S.G., Huppert, T.J., Kolehmainen, V., Franceschini, M.A., Kaipio, J.P., Arridge, S.R., Boas, D.A., 2006. Dynamic physiological modeling for functional diffuse optical tomography. *Neuroimage* 30, 88–101.

Fischl, B., Sereno, M.I., Dale, A.M., 1999. Cortical surface-based analysis: II: Inflation, flattening, and a surface-based coordinate system. *Neuroimage* 9, 195–207.

Franceschini, M.A., Boas, D.A., 2004. Noninvasive measurement of neuronal activity with near-infrared optical imaging. *Neuroimage* 21, 372–386.

Franceschini, M.A., Joseph, D.K., Huppert, T.J., Diamond, S.G., Boas, D.A., 2006. Diffuse optical imaging of the whole head. *J. Biomed. Opt.* 11, 054007.

Friston, K.J., Zarahn, E., Josephs, O., Henson, R.N.A., Dale, A.M., 1999. Stochastic designs in event-related fMRI. *Neuroimage* 10, 607–619.

Friston, K.J., Mechelli, A., Turner, R., Price, C.J., 2000. Nonlinear responses in fMRI: the Balloon model, Volterra kernels, and other hemodynamics. *Neuroimage* 12, 466–477.

Gibson, A.P., Hebdon, J.C., Arridge, S.R., 2005. Recent advances in diffuse optical imaging. *Phys. Med. Biol.* 50, R1–R43.

Glover, G.H., Li, T.-Q., Ress, D., 2000. Image-based method for retrospective correction of physiological motion effects in fMRI: RETROICOR. *Magn. Reson. Med.* 44, 162–167.

Greve, D.N., Fischl, B., 2009. Accurate and robust brain image alignment using boundary-based registration. *Neuroimage* 48, 63–72.

Greve, D.N., Goldenholz, D., Kaskhedikar, G., Polimeni, J.R., 2009. BOLD Physiological Noise Reduction using Spatio-Spectral-Temporal Correlations with NIRS ISMRM, Hawaii (2009). ISMRM, Hawaii.

Greve, D.N., Mueller, B.A., Liu, T., Turner, J.A., Voyvodic, J., Yetter, E., Diaz, M., McCarthy, G., Wallace, S., Roach, B.J., Ford, J.M., Mathalon, D.H., Calhoun, V.D., Wible, C.G., Brown, G.G., Potkin, S.G., Glover, G., 2011. A novel method for quantifying scanner instability in fMRI. *Magn. Reson. Med.* 65, 1053–1061.

- Gustafsson, H., 1993. Vasomotion and underlying mechanisms in small arteries. An in vitro study of rat blood vessels. *Acta. Physiol. Scand. (Suppl. 614)*, 1–44.
- Hu, X., Le, T.H., Parrish, T., Erhard, P., 1995. Retrospective estimation and correction of physiological fluctuation in functional MRI. *Magn. Reson. Med.* 34, 201–212.
- Huettel, S., Song, A., McCarthy, G., 2004. *Functional Magnetic Resonance Imaging*. Sinauer Associates.
- Huppert, T.J., Hoge, R.D., Diamond, S.G., Franceschini, M.A., Boas, D.A., 2006. A temporal comparison of BOLD, ASL, and NIRS hemodynamic responses to motor stimuli in adult humans. *Neuroimage* 29, 368–382.
- Huppert, T.J., Allen, M.S., Diamond, S.G., Boas, D.A., 2009. Estimating cerebral oxygen metabolism from fMRI with a dynamic multicompartment Windkessel model. *Hum. Brain Mapp.* 30, 1548–1567.
- Jones, T.B., Bandettini, P.A., Birn, R.M., 2008. Integration of motion correction and physiological noise regression in fMRI. *Neuroimage* 42, 582–590.
- Julien, C., 2006. The enigma of Mayer waves: facts and models. *Cardiovasc. Res.* 70, 12–21.
- Lina, J.-M., Dehaes, M., Matteau-Pelletier, C., Lesage, F., 2008. Complex wavelets applied to diffuse optical spectroscopy for brain activity detection. *Opt. Express* 16, 1029–1050.
- Obrig, H., Villringer, A., 2003. Beyond the visible—imaging the human brain with light. *J. Cereb. Blood Flow Metab.* 23, 1–18.
- Prince, S., Kolehmainen, V., Kaipio, J.P., Franceschini, M.A., Boas, D., Arridge, S.R., 2003. Time-series estimation of biological factors in optical diffusion tomography. *Phys. Med. Biol.* 48, 1491–1504.
- Smith, S.M., 2002. Fast robust automated brain extraction. *Hum. Brain Mapp.* 17, 143–155.
- Steinbrink, J., Villringer, A., Kempf, F., Haux, D., Boden, S., Obrig, H., 2006. Illuminating the BOLD signal: combined fMRI–fNIRS studies. *Magn. Reson. Imaging* 24, 495–505.
- Strangman, G., Culver, J.P., Thompson, J.H., Boas, D.A., 2002. A quantitative comparison of simultaneous BOLD fMRI and NIRS recordings during functional brain activation. *Neuroimage* 17, 719–731.
- Tong, Y., Frederick, B.D., 2010. Time lag dependent multimodal processing of concurrent fMRI and near-infrared spectroscopy (NIRS) data suggests a global circulatory origin for low-frequency oscillation signals in human brain. *Neuroimage* 53, 553–564.
- Tong, Y., Bergethon, P.R., deB, Frederick, B., 2011. An improved method for mapping cerebrovascular reserve using concurrent fMRI and near-infrared spectroscopy with Regressor Interpolation at Progressive Time Delays (RIPTiDe). *Neuroimage* 56, 2047–2057.
- Toronov, V., Webb, A., Choi, J.H., Wolf, M., Michalos, A., Gratton, E., Hueber, D., 2001. Investigation of human brain hemodynamics by simultaneous near-infrared spectroscopy and functional magnetic resonance imaging. *Med. Phys.* 28, 521.
- Villringer, A., Planck, J., Hock, C., Schleinkofer, L., Dirnagl, U., 1993. Near infrared spectroscopy (NIRS): a new tool to study hemodynamic changes during activation of brain function in human adults. *Neurosci. Lett.* 154, 101–104.
- Wise, R.G., Ide, K., Poulin, M.J., Tracey, I., 2004. Resting fluctuations in arterial carbon dioxide induce significant low frequency variations in BOLD signal. *Neuroimage* 21, 1652–1664.